



Protocol for the Examination of Resection Specimens From Patients With Primary Tumors of Bone

Version: 4.2.0.0

Protocol Posting Date: June 2024

CAP Laboratory Accreditation Program Protocol Required Use Date: March 2025

The changes included in this current protocol version affect accreditation requirements. The new deadline for implementing this protocol version is reflected in the above accreditation date.

For accreditation purposes, this protocol should be used for the following procedures and tumor types:

Procedure	Description
Resection	Includes specimens designated intralesional resection, marginal resection, segmental resection / limb salvage, wide resection, or radical resection / amputation.
Tumor Type	Description
Primary malignant bone tumors	Includes tumors arising in bone for which pTNM staging is clinically relevant.

This protocol is **NOT** required for accreditation purposes for the following:

Procedure
Biopsy (includes core, curettage, or incisional, consider Bone Biopsy Protocol)
Primary resection specimen with no residual / viable cancer (e.g., following neoadjuvant therapy)
Cytologic fine needle aspiration (FNA) specimens
Tumor Type
Bone tumors that may recur locally but have either no or an extremely low risk of metastasis

The following tumor types should **NOT** be reported using this protocol:

Tumor Type
Lymphoma / Leukemia (consider the Precursor and Mature Lymphoid Malignancies, Myeloid and Mixed / Ambiguous Lineage Neoplasms, or Plasma Cell Malignancies protocols)
Pediatric Ewing sarcoma (consider the Pediatric Ewing Sarcoma protocol)
Soft tissue primary sarcoma (consider the Soft Tissue protocol)

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With guidance from the CAP Cancer and CAP Pathology Electronic Reporting Committees.

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Accreditation Requirements

This protocol can be utilized for a variety of procedures and tumor types for clinical care purposes. For accreditation purposes, only the definitive primary cancer resection specimen is required to have the core and conditional data elements reported in a synoptic format.

- Core data elements are required in reports to adequately describe appropriate malignancies. For accreditation purposes, essential data elements must be reported in all instances, even if the response is “not applicable” or “cannot be determined.”
- Conditional data elements are only required to be reported if applicable as delineated in the protocol. For instance, the total number of lymph nodes examined must be reported, but only if nodes are present in the specimen.
- Optional data elements are identified with “+” and although not required for CAP accreditation purposes, may be considered for reporting as determined by local practice standards.

The use of this protocol is not required for recurrent tumors or for metastatic tumors that are resected at a different time than the primary tumor. Use of this protocol is also not required for pathology reviews performed at a second institution (i.e., secondary consultation, second opinion, or review of outside case at second institution).

Synoptic Reporting

All core and conditionally required data elements outlined on the surgical case summary from this cancer protocol must be displayed in synoptic report format. Synoptic format is defined as:

- Data element: followed by its answer (response), outline format without the paired Data element: Response format is NOT considered synoptic.
- The data element should be represented in the report as it is listed in the case summary. The response for any data element may be modified from those listed in the case summary, including “Cannot be determined” if appropriate.
- Each diagnostic parameter pair (Data element: Response) is listed on a separate line or in a tabular format to achieve visual separation. The following exceptions are allowed to be listed on one line:
 - Anatomic site or specimen, laterality, and procedure
 - Pathologic Stage Classification (pTNM) elements
 - Negative margins, as long as all negative margins are specifically enumerated where applicable
- The synoptic portion of the report can appear in the diagnosis section of the pathology report, at the end of the report or in a separate section, but all Data element: Responses must be listed together in one location

Organizations and pathologists may choose to list the required elements in any order, use additional methods in order to enhance or achieve visual separation, or add optional items within the synoptic report. The report may have required elements in a summary format elsewhere in the report IN ADDITION TO but not as replacement for the synoptic report i.e., all required elements must be in the synoptic portion of the report in the format defined above.

Summary of Changes

v 4.2.0.0

- Cover page update
- Updates to content and explanatory notes, including WHO Histologic Types
- pTNM Classification update
- LVI question update from optional to required (core) and “Lymphovascular Invasion” to “Lymphatic and / or Vascular Invasion”
- “Other Close Margin(s) to Tumor” criterion update
- “Mitotic Rate” answer update
- Addition of required (core) questions “Treatment Effect (for post-neoadjuvant therapy)”, and “Tumor Laterality”
- Updates to conditional question “Necrosis in the Absence of Neoadjuvant therapy (required only if neoadjuvant therapy was not administered)”
- Addition of optional questions “Associated Syndrome”, “Other Clinical Findings”, and “Decalcification Procedure”
- SPECIAL STUDIES section update

Reporting Template

Protocol Posting Date: June 2024

Select a single response unless otherwise indicated.

CASE SUMMARY: (BONE: Resection)

Standard(s): AJCC-UICC 8

CLINICAL (Note [A](#))

+Associated Syndrome

- Li-Fraumeni syndrome
- Mazabraud syndrome
- Ollier disease
- Maffucci syndrome
- Hereditary multiple exostoses
- Other (specify): _____
- Not specified

+Radiologic Findings (Notes [A](#),[B](#))

- Specify: _____
- Not available

+Preresection Treatment (select all that apply)

- No known preresection therapy
- Chemotherapy
- Radiation therapy
- Other (specify): _____
- Therapy administered, type not specified
- Not specified

+Other Clinical Findings

- Specify: _____
- Not available

SPECIMEN

Procedure (Note [C](#))

- Intralesional resection
- Marginal resection
- Segmental / wide resection
- Limb salvage (specify): _____
- Radical resection of bone and soft tissue
- Amputation (specify): _____
- Other (specify): _____
- Not specified

+Decalcification Procedure (Note C)

- EDTA-decal or equivalent
- Harsh acid decalcification

TUMOR

Multiple Sites (required only if applicable)

- Not applicable
- Multifocal tumor / discontinuous tumor at primary bone site
- Additional primary bone site(s) present (specify for synchronous malignant tumors or polyostotic aggressive tumors) : _____

Tumor Site (Note D)

- Appendicular skeleton, trunk, skull, facial bones (specify): _____
- Spine (specify bone, if known): _____
- Pelvis (specify bone, if known): _____
- Not specified

Tumor Laterality

- Left
- Right
- Central
- Polyostotic ipsilateral
- Polyostotic bilateral
- Cannot be determined

Tumor Location and Extent (Note B) (select all that apply)

- Epiphysis or apophysis
- Metaphysis
- Diaphysis
- Cortex
- Medullary cavity
- Surface
- Involves joint
- Extends into soft tissue
- Cannot be determined: _____

Tumor Size

- Greatest dimension in Centimeters (cm): _____ cm
- +Additional Dimension in Centimeters (cm): _____ x _____ cm**
- +Radiologic Greatest Dimension in Centimeters (cm): _____ cm**
- Cannot be determined: _____

Histologic Type# (Note E)

The list is derived from the World Health Organization (WHO) classification of bone tumors, 5th edition, to include ONLY bone tumors of intermediate (locally aggressive and rarely metastasizing) potential and malignant bone tumors.

- ___ Chondrogenic tumors
 - ___ Synovial chondromatosis
 - ___ Atypical cartilaginous tumor
 - ___ Chondrosarcoma
 - ___ Chondrosarcoma, secondary (specify): _____
 - ___ Dedifferentiated chondrosarcoma
 - ___ Periosteal chondrosarcoma
 - ___ Clear cell chondrosarcoma
 - ___ Mesenchymal chondrosarcoma
- ___ Osteogenic tumors
 - ___ Osteoblastoma
 - ___ Low-grade central osteosarcoma
 - ___ Low-grade central osteosarcoma with high-grade transformation
 - ___ Parosteal osteosarcoma
 - ___ Parosteal osteosarcoma with high-grade transformation
 - ___ Conventional osteosarcoma
 - ___ Telangiectatic osteosarcoma
 - ___ Small cell osteosarcoma
 - ___ Periosteal osteosarcoma
 - ___ High-grade surface osteosarcoma
 - ___ Secondary osteosarcoma
 - ___ **+Precipitating Factor for Secondary Osteosarcoma:** _____
- ___ Undifferentiated small round cell sarcomas
 - ___ Ewing sarcoma
 - ___ Round cell sarcoma with EWSR1::non-ETS fusions (specify, if known): _____
 - ___ CIC-rearranged sarcoma
 - ___ Sarcoma with BCOR genetic alterations
- ___ Fibrogenic / fibrohistiocytic / histiocytic tumors
 - ___ Sclerosing epithelioid fibrosarcoma
 - ___ Primary malignant giant cell tumor of bone
 - ___ Secondary malignant giant cell tumor of bone
 - ___ Giant cell tumor of bone
 - ___ Langerhans cell histiocytosis
 - ___ **+System Involvement**
 - ___ Single system (specify): _____
 - ___ Multisystem (specify): _____
 - ___ Other (leukemic, atypical, or other, specify): _____
- ___ Desmoplastic fibroma
- ___ Notochordal tumors
 - ___ Conventional chordoma
 - ___ Poorly differentiated chordoma
 - ___ Dedifferentiated chordoma
- ___ Vascular tumors

- Epithelioid hemangioma
 - Pseudomyogenic hemangioendothelioma
 - Epithelioid hemangioendothelioma
 - Angiosarcoma
 - Epithelial tumors
 - Adamantinoma of long bones
 - Osteofibrous dysplasia-like adamantinoma
 - Dedifferentiated adamantinoma
 - Other mesenchymal tumors or tumors of uncertain differentiation
 - Leiomyosarcoma of bone
 - Rhabdomyosarcoma of bone (specify fusion, if known): _____
 - TK-fusion (NTRK, ALK, BRAF) tumor, primary intraosseous (specify fusion, if known): _____
 - Undifferentiated pleomorphic sarcoma
 - Cannot be determined: _____
 - Other histologic type not listed (specify): _____
- +Histologic Type Comment:** _____

Histologic Grade (Note F)

- G1, well-differentiated, low-grade
- G2, moderately differentiated, high-grade
- G3, poorly differentiated, high-grade
- GX, cannot be assessed: _____
- Ungraded tumor / not applicable for this tumor type

+Mitotic Rate (Note G)

- Specify mitotic rate per mm²: _____ mitoses per mm²
- Specify mitotic rate per 10 high-power fields (HPF): _____ mitoses per 10 high-power fields (HPF)
- Cannot be determined (explain): _____

Treatment Effect (for post-neoadjuvant treatment) (Note H)

- No known presurgical therapy
 - Not identified
- # Therapy response is expressed as a percentage of total tumor area that is non-viable.*
- Present (specify overall percentage of treatment effect)#: _____ %
- Select all that apply*
- + Geographic necrosis
 - + Fibrosis
 - + Hyalinization
 - + Hemorrhage
 - + Cystic change
 - + Histiocytic response
 - + Inflammation
 - + Other (specify): _____
 - Cannot be determined

Necrosis in the Absence of Neoadjuvant Therapy (required only if neoadjuvant therapy was not administered)

- Not applicable (neoadjuvant therapy was administered)
- Not identified
- Present

Extent of Necrosis

- Specify percentage: _____ %
- Other (specify): _____
- Cannot be determined (explain): _____
- Cannot be determined

Lymphatic and / or Vascular Invasion (Note I)

- Not identified
- Present
- Cannot be determined

+Tumor Comment: _____

MARGINS (Note J)

Margin Status

- All margins negative for tumor

Closest Margin(s) to Tumor

- Specify closest margin(s): _____
- Cannot be determined (explain): _____

Distance from Tumor to Closest Margin

Specify in Centimeters (cm)

- Exact distance: _____ cm
- Greater than: _____ cm
- At least: _____ cm
- Less than: _____ cm
- Other (specify): _____
- Cannot be determined: _____

+Other Close Margin(s) to Tumor (less than 0.2 cm)

- Specify other close margin(s): _____
- Cannot be determined (explain): _____
- Not applicable
- Tumor present at margin

Margin(s) Involved by Tumor

- Specify involved margin(s): _____
- Cannot be determined: _____
- Other (specify): _____
- Cannot be determined (explain): _____
- Not applicable

+Margin Comment: _____

REGIONAL LYMPH NODES (Note [K](#))

Regional Lymph Node Status

- Not applicable (no regional lymph nodes submitted or found)
- Regional lymph nodes present
 - All regional lymph nodes negative for tumor
 - Tumor present in regional lymph node(s)

Number of Lymph Nodes with Tumor

- Exact number (specify): _____
- At least (specify): _____
- Other (specify): _____
- Cannot be determined (explain): _____
- Other (specify): _____
- Cannot be determined (explain): _____

Number of Lymph Nodes Examined

- Exact number (specify): _____
- At least (specify): _____
- Other (specify): _____
- Cannot be determined (explain): _____

+Regional Lymph Node Comment: _____

DISTANT METASTASIS

Distant Site(s) Involved, if applicable (select all that apply)

- Not applicable
- Lung: _____
- Bone: _____
- Other (specify): _____
- Cannot be determined: _____

pTNM CLASSIFICATION (AJCC 8th Edition) (Note [L](#))

Reporting of pT, pN, and (when applicable) pM categories is based on information available to the pathologist at the time the report is issued. As per the AJCC (Chapter 1, 8th Ed.) it is the managing physician's responsibility to establish the final pathologic stage based upon all pertinent information, including but potentially not limited to this pathology report.

pTNM Classification (required only if applicable)

Regardless of the anatomic site, certain specific types of bone neoplasms for which pTNM staging is not clinically relevant are excluded from the staging system.

- Not applicable (histologic type not appropriate for staging)#
- Histologic type appropriate for staging

Modified Classification (required only if applicable) (select all that apply)

- Not applicable
- y (post-neoadjuvant therapy)
- r (recurrence)

pT Category

___ Appendicular skeleton, trunk, skull, and facial bones

pT Category

___ pT not assigned (cannot be determined based on available pathological information)

___ pT0: No evidence of primary tumor

___ pT1: Tumor less than or equal to 8 cm in greatest dimension

___ pT2: Tumor greater than 8 cm in greatest dimension

___ pT3: Discontinuous tumors in the primary bone site

___ Spine

pT Category

___ pT not assigned (cannot be determined based on available pathological information)

___ pT0: No evidence of primary tumor

___ pT1: Tumor confined to one vertebral segment or two adjacent vertebral segments

___ pT2: Tumor confined to three adjacent vertebral segments

___ pT3: Tumor confined to four or more adjacent vertebral segments, or any nonadjacent vertebral segments

pT4: Extension into the spinal canal or great vessels

___ pT4a: Extension into the spinal canal

___ pT4b: Evidence of gross vascular invasion or tumor thrombus in the great vessels

___ pT4 (subcategory cannot be determined)

___ Pelvis

pT Category

___ pT not assigned (cannot be determined based on available pathological information)

___ pT0: No evidence of primary tumor

pT1: Tumor confined to one pelvic segment with no extraosseous extension

___ pT1a: Tumor less than or equal to 8 cm in greatest dimension

___ pT1b: Tumor greater than 8 cm in greatest dimension

___ pT1 (subcategory cannot be determined)

pT2: Tumor confined to one pelvic segment with extraosseous extension or two segments without extraosseous extension

___ pT2a: Tumor less than or equal to 8 cm in greatest dimension

___ pT2b: Tumor greater than 8 cm in greatest dimension

___ pT2 (subcategory cannot be determined)

pT3: Tumor spanning two pelvic segments with extraosseous extension

___ pT3a: Tumor less than or equal to 8 cm in greatest dimension

___ pT3b: Tumor greater than 8 cm in greatest dimension

___ pT3 (subcategory cannot be determined)

pT4: Tumor spanning three pelvic segments or crossing the sacroiliac joint

___ pT4a: Tumor involves sacroiliac joint and extends medial to the sacral neuroforamen

___ pT4b: Tumor encasement of external iliac vessels or presence of gross tumor thrombus in major pelvic vessels

___ pT4 (subcategory cannot be determined)

T Suffix (required only if applicable)

___ Not applicable

___ (m) multiple primary synchronous tumors in single anatomic site

pN Category (Note K)

___ pN not assigned (no nodes submitted or found)

___ pN not assigned (cannot be determined based on available pathological information)

___ pN0: No regional lymph node metastasis

___ pN1: Regional lymph node metastasis

pM Category (required only if confirmed pathologically)

___ Not applicable - pM cannot be determined from the submitted specimen(s)

pM1: Distant metastasis

___ pM1a: Lung

___ pM1b: Bone or other distant sites

___ pM1 (subcategory cannot be determined)

ADDITIONAL FINDINGS

+Additional Findings (specify): _____

SPECIAL STUDIES (Note E)

The previously reported biopsy immunohistochemistry, cytogenetics, and molecular studies can be included in the resection report.

Immunohistochemistry

___ Specify results: _____

___ Pending (specify): _____

___ Not performed: _____

___ Not applicable

___ Other (specify): _____

Cytogenetics

___ Specify results: _____

___ Pending (specify): _____

___ Not performed: _____

___ Not applicable

___ Other (specify): _____

Molecular Studies

___ Specify results: _____

___ Pending (specify): _____

___ Not performed: _____

___ Not applicable

___ Other (specify): _____

COMMENTS

Comment(s): _____

Explanatory Notes

A. Scope of Guidelines

Anatomical staging using the AJCC system 8th ed. is considered clinically relevant only for the specific malignant entities listed in World Health Organization classification 5th ed. This includes malignant chondrogenic tumors, osteogenic tumors, fibrogenic tumors, osteoclastic giant cell-rich tumors, notochordal tumors, myogenic tumors, lipogenic tumors, undifferentiated small round cell sarcomas and other mesenchymal tumors arising in bone. Locally aggressive entities such as synovial chondromatosis, osteoblastoma, giant cell tumor of bone, epithelioid hemangioma, pseudomyogenic hemangioma, and desmoplastic fibroma may be reported using this protocol but are not staged. Radiologic parameters include bone involved, size and extent (compartment) of tumor, location of tumor and extent, radiologic intrinsic characteristics including matrix or mineralization in bone-forming tumors, and differential diagnosis. Clinical parameters include patient age, sex, exact anatomic location, size, solitary or polyostotic, syndromes, and other pertinent medical and surgical history, if clinically relevant.

B. Tumor Location and Extent

Radiographic imaging plays an especially critical role in the diagnosis of bone tumors. Close collaboration with an experienced musculoskeletal radiologist and orthopedic surgeon is advised.

Figure 1 is a diagrammatic representation of the “anatomic” regions of a long bone. These locations are very important in classifying bone tumors. For instance, chondroblastoma almost always arises in the epiphysis. Epiphyses and apophyses are secondary ossification centers and therefore are embryonic equivalents; “epiphyses” are found within joints, whereas “apophyses”, the sites of tendonous and ligamentous attachments, are not found within joints. The greater and lesser trochanters are apophyses, while the epiphyses are at the ends of long bones.

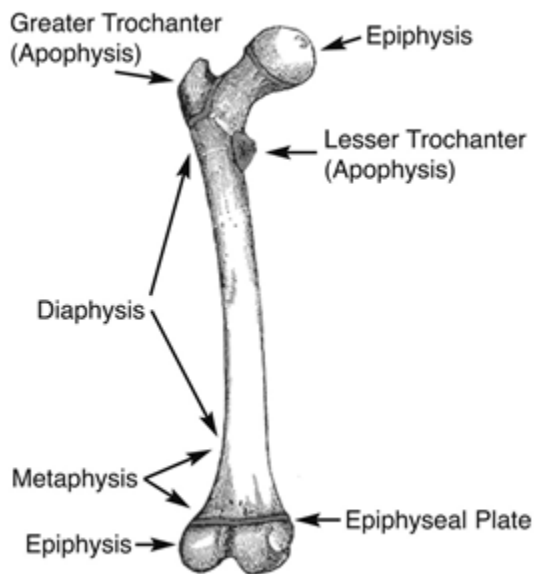


Figure 1. Important anatomic landmarks for tumor diagnosis in long bones. Adapted from Gray's Anatomy.¹

References

1. Gray H, Lewis WH. *Gray's Anatomy of the Human Body*. 20th ed. Philadelphia, PA: Lea & Febiger; 1918.

C. Procedure/Tissue Processing/Tissue for Genetic-Molecular Studies

The following is a list of guidelines to be used in defining what type of procedure has been performed. This is based on the surgeon's intent and not based on the pathologic assessment of the margins.

Intralesional Resection: Leaving gross tumor behind. Partial debulking is an example.

Marginal Resection: Removing the tumor and its pseudocapsule with a relatively small amount of adjacent tissue. There is no gross tumor at the margin; however, microscopic tumor may be present. Note that occasionally, a surgeon will perform an "excisional" biopsy, which effectively accomplishes the same thing as a marginal resection.

Segmental/Wide Resection: An intracompartmental resection. A single piece of bone is resected, including the lesion, adjacent soft tissue, and a cuff of normal bone. Limb salvage is an example.

Radical Resection: The removal of an entire bone, and the excision of the adjacent muscle groups if the tumor is extracompartmental. Amputation is an example.

Fixation

Tissue specimens from bone tumors optimally are received fresh/unfixed in case fresh tissue for ancillary studies, such as cytogenetics, is required. All tissue should be processed in a manner that would allow molecular studies to be undertaken successfully.^{1,2,3} Decalcification using harsh acidic reagents may be detrimental for nucleic acid-based molecular studies and therefore utilization of EDTA as a decalcifying agent has been advised. Freezing a portion of the sample and/or fixing soft portions of the lesion in buffered formalin is encouraged over EDTA decalcification, for molecular studies.

Tissue Submission for Histologic Evaluation and Genetic/Molecular Studies

One section per centimeter of maximum dimension is usually recommended, although fewer sections are needed for very large tumors, especially if these are homogeneous. Tumors known to be high grade from a previous biopsy do not require as many sections as those that were previously diagnosed as low grade, as documentation of a high-grade component will change stage, prognosis, and treatment in the latter case. All bone tumors that are post-adjuvant therapy (chemotherapy or radiation) have a central slab section entirely mapped and submitted to assess necrosis. Cystic or hemorrhagic areas should be grossly estimated, and the rim entirely submitted. Always, sections should be taken of grossly heterogeneous areas.^{4,5} Occasionally, gross findings can be misleading, and areas that appear to be necrotic may actually be myxoid or edematous. Tumors that have not had pre-adjuvant chemotherapy require at least 1 section per cm and all heterogeneous areas, including those appearing necrotic.

While it has been helpful and often required for clinical trials to have snap frozen tissue, approximately 1 cm³ of fresh tissue stored at minus seventy (-70° C) that can be shipped on dry ice to facilities that perform molecular analysis, most full evaluations of sarcomas can be made on formalin-fixed and EDTA decalcified paraffin-embedded tissue. Adequate tissue should be submitted for conventional light microscopy before tissue has been taken for other analysis. Other than for flow cytometry, microbiology

cultures (to send directly to the Microbiology Laboratory from the Operating Room), EM, and cytogenetics, most additional studies for FISH, molecular and copy number/methylation profiling can all be performed on both formalin-fixed and EDTA-fixed tissue (and not on acid decalcified tissue).

Intraoperative Consultation

Most intraoperative assessment is for margins. Histologic classification of bone tumors is sufficiently complex that it is unreasonable to expect a precise classification of these tumors based on an intraoperative consultation. A complete understanding of the surgeon's treatment algorithm is recommended before rendering a frozen section diagnosis. In the case of primary bone tumors, an intraoperative diagnosis of benign versus malignant will generally guide the immediate decision to curette, excise, or wait for permanent sections, and certain therapeutic options may be lost if the wrong path is pursued. Intraoperative consultation is useful in assessing if "lesional" tissue is present and whether or not this tissue is necrotic, and in constructing a differential diagnosis that can direct the proper triage of tissue for flow cytometry (lymphoma), electron microscopy, and molecular studies/cytogenetics. Tissue triage optimally is performed at the time of frozen section.

References

1. Taylor BS, Barretina J, Maki RG, Antonescu CR, Singer S, Ladanyi M. Advances in sarcoma genomics and new therapeutic targets. *Nat Rev Cancer*. 2011;11(8):541-547.
2. Rubin BP, Lazar JF, Oliveira AM. Molecular pathology of bone and soft tissue tumors. In: Tubbs R, Stoler M. *Cell and Tissue Based Molecular Pathology*. Philadelphia, PA: Churchill Livingstone; 2009.
3. WHO Classification of Tumours Editorial Board. Soft Tissue and Bone Tumors. Lyon (France): International Agency for Research on Cancer; 2020. (WHO classification of tumours series, 5th ed.; vol. 3).
4. Amin MB, Edge SB, Greene FL, et al., eds. *AJCC Cancer Staging Manual*. 8th ed. New York, NY: Springer; 2017.
5. Pawel B, Bahrami A, Hicks MJ, Rudzinski E. Protocol for the Examination of Specimens From Pediatric Patients With Ewing Sarcoma (ES). 2016. Available at www.cap.org/cancerprotocols.

D. Tumor Site

Given the strong association between the primary anatomic site and outcome, the 8th edition of the AJCC Cancer Staging Manual⁴ uses the following site groups for staging purposes:

- Appendicular skeleton, including trunk, skull, and facial bones
- Pelvis
- Spine

This site grouping is reflected by the provision of separate definitions for the primary tumor (T) for each anatomic site.

References

1. Amin MB, Edge SB, Greene FL, et al., eds. *AJCC Cancer Staging Manual*. 8th ed. New York, NY: Springer; 2017.

E. Classification of Bone Tumors

The list is derived from the World Health Organization (WHO) classification of soft tissue tumors, 5th edition,¹ edited to include ONLY bone tumors of intermediate (locally aggressive and rarely metastasizing) potential and malignant bone tumors.

Note on atypical cartilaginous tumor/grade 1 chondrosarcoma:

Atypical cartilaginous tumor (ACT) refers to cartilaginous neoplasms demonstrating features of a grade 1 chondrosarcoma and arising in the short and long tubular bones. This terminology should not be used when a pathologist cannot decide on the classification for the cartilaginous neoplasm.

Bone Primary Tyrosine Kinase Fusion Tumors:

While fusions involving the *RAS::MAPK* pathway are rare among bone tumors, these tumors have driver alterations in genes that encode tyrosine kinases and may respond to therapy targeting *NTRK*, *ALK*, *BRAF*, *RET*, *RAF*, *FGFR1*, or *ABL1*, etc. Notably, *NTRK* tumors fused with *KANK1* or *TPR* have been demonstrated to exhibit higher-grade appearance, including spindled and pleomorphic characteristics, accompanied by necrosis and mitoses, leading to unfavorable outcomes. Consequently, it is advisable to conduct comprehensive RNA-based Next-Generation Sequencing (NGS) for fusions, particularly in spindled pleomorphic tumors occurring in individuals under 50 years old, especially those in soft tissue or intraosseous locations. This recommendation is especially pertinent with tumors that have variable ovoid spindled to epithelioid morphology, variable collagenous to myxoid stroma, variable gaping to staghorn vasculature and specifically focal CD34 and/or focal S100 protein, without any staining for SOX10. In these tumors, BRAF, ALK, or panTrk or other immunostain may be identified.^{2,3,4,5,6,7,8,9,10,11}

Most Common Molecular/Genetic Findings:

The most common molecular/genetic findings in a subset of intermediate/malignant bone tumors are listed (Table 1).

Table 1: Subset of bone intermediate and malignant tumors with the most common diagnostic molecular/genetic findings.

<u>Diagnosis</u>	<u>Genes Involved</u>
Chondrosarcoma	<i>IDH1/IDH2</i> mutation
Intraosseous extraskeletal myxoid chondrosarcoma	<i>EWSR1/TAF15::NR4A3</i> fusion
Mesenchymal chondrosarcoma	<i>HEY1::NCOA2</i> fusion
Secondary chondrosarcoma arising in enchondroma	<i>IDH1/IDH2</i> mutation
Secondary chondrosarcoma arising in osteochondroma	<i>EXT1/EXT2</i> mutation
Sclerosing epithelioid fibrosarcoma of bone	<i>FUS::CREB3L2</i> fusion
Angiomatoid fibrous histiocytoma of bone/joint	<i>EWSR1::CREB1</i> or <i>EWSR1::ATF1</i> alternate

Primary malignant giant cell tumor of bone	<i>H3F3A</i> mutation
Leukemia/Multifocal atypical Langerhans cell histiocytosis	<i>BRAF</i> mutation
Poorly differentiated chordoma	<i>SMARCB1</i> deletion
Low-grade central osteosarcoma	<i>MDM2/CDK4</i> amplification
Parosteal osteosarcoma	<i>MDM2/CDK4</i> amplification
Rhabdomyosarcoma of bone (adult)	<i>FUS/EWSR1::TFCP2</i> , <i>MEIS1::NCOA2</i>
Ewing sarcoma	<i>EWSR1::FLI1</i> (85-90%), <i>EWSR1::ERG</i> (8-10%), others
<i>CIC</i> -rearranged sarcoma	<i>CIC::DUX4</i>
Round cell sarcoma with <i>EWSR1::non-ETS</i> fusion	<i>EWSR1::PATZ1</i> , <i>EWSR1::NFATC2</i> , <i>FUS::NFATC2</i>
Sarcoma with <i>BCOR</i> genetic alterations	<i>BCOR::CCNB3</i> fusion
Epithelioid hemangioendothelioma of bone	<i>WWTR1::CAMTA1</i> fusion
Angiosarcoma of bone	<i>MYC</i> amplification (post-irradiation)
Tyrosine-kinase fusion tumor	<i>NTRK1/2/3</i> , <i>ALK</i> , <i>BRAF</i> , etc. fusion (various fusion partners)

References

1. WHO Classification of Tumours Editorial Board. Soft Tissue and Bone Tumors. Lyon (France): International Agency for Research on Cancer; 2020. (WHO classification of tumours series, 5th ed.; vol. 3)
2. Haller F, Knopf J, Ackermann A, et al. Paediatric and adult soft tissue sarcomas with NTRK1 gene fusions: a subset of spindle cell sarcomas unified by a prominent myopericytic/haemangiopericytic pattern. *J Pathol*. 2016 Apr;238(5):700-10.
3. Hung YP, Fletcher CDM, Hornick JL. Evaluation of pan-TRK immunohistochemistry in infantile fibrosarcoma, lipofibromatosis-like neural tumour and histological mimics. *Histopathology*. 2018;73(4):634-644.
4. Agaram NP, Zhang L, Sung YS, et al. Recurrent NTRK1 Gene Fusions Define a Novel Subset of Locally Aggressive Lipofibromatosis-like Neural Tumors. *Am J Surg Pathol*. 2016 Oct;40(10):1407-16.
5. Helm M, Chang A, Fanburg-Smith JC, Zaenglein AL, Helm K. Cutaneous VCL::ALK fusion ovoid-spindle cell neoplasm. *J Cutan Pathol*. 2023;50(5):405-409. doi: 10.1111/cup.14420. Epub 2023 Mar 12. PMID: 36843055.
6. Fanburg-Smith JC, Smith JD, Flemming DJ. Bone and soft tissue tumors: clinicoradiologic-pathologic molecular-genetic correlation of novel fusion spindled, targetable-ovoid, giant-cell-rich, and round cell sarcomas. *Skeletal Radiol*. 2023 Mar;52(3):517-540. doi: 10.1007/s00256-022-04244-w. Epub 2022 Dec 21. PMID: 36542130.

7. Wood ML, Fanburg-Smith JC, Brian JM, White JC, Powell JL, Freiberg AS. Successful Crizotinib-targeted Therapy of Pediatric Unresectable ERC1::ALK Fusion Sarcoma. *J Pediatr Hematol Oncol*. 2023. doi: 10.1097/MPH.0000000000002777. Epub ahead of print. PMID: 38099690.
8. Davis JL, Lockwood CM, Stohr B, et al. Expanding the Spectrum of Pediatric NTRK-rearranged Mesenchymal Tumors. *Am J Surg Pathol*. 2019 Apr;43(4):435-445.
9. Chen T, Wang Y, Goetz L, Corey Z, Dougher MC, Smith JD, Fox EJ, Freiberg AS, Flemming D, Fanburg-Smith JC. Novel fusion sarcomas including targetable NTRK and ALK. *Ann Diagn Pathol*. 2021;54:151800. PMID: 34464935.
10. Tan SY, Al-Ibraheemi A, Ahrens WA, Oesterheld JE, Fanburg-Smith JC, Liu YJ, Spunt SL, Rudzinski ER, Coffin C, Davis JL. ALK rearrangements in infantile fibrosarcoma-like spindle cell tumours of soft tissue and kidney. *Histopathology*. 2022 Mar;80(4):698-707. Epub 2022 Jan 2. PMID: 34843129.
11. Eyerer FIR, Bradshaw G, Vasalos P, Laser JS, Chang CC, Kim AS, Olson DR, Paler RJ, Rosenbaum JN, Walk EE, Willis JE, Yao J, Yohe SL. Getting Your Laboratory on Track With Neurotrophic Receptor Tyrosine Kinase. *Arch Pathol Lab Med*. 2023 Aug 1;147(8):872-884.

F. Grading

The grading of bone tumors is largely driven by the histologic diagnosis, and traditionally grading has been based on the system advocated by Broders, which assesses cellularity and nuclear features/degree of anaplasia.¹ The eighth edition of the AJCC Cancer Staging Manual recommends a 2-tiered system (low vs high-grade) for recording grading.² Histologic grading uses a 3-tiered system: Grade 1 is considered low-grade, and Grade 2 and Grade 3 are grouped together as high-grade for biological grading. In bone sarcomas, the histologic subtype often determines the clinical behavior and grade. Therefore, a more pragmatic approach to grading aggressive and malignant primary tumors of bone can be used.³

Two bone tumors that are locally aggressive and metastasize infrequently, and thus are usually low-grade, are low-grade central osteosarcoma and parosteal osteosarcoma. Periosteal osteosarcoma is generally regarded as a grade 2 osteosarcoma. Primary bone tumors that are generally high-grade include malignant giant cell tumor, Ewing sarcoma, angiosarcoma, dedifferentiated chondrosarcoma, conventional osteosarcoma, telangiectatic osteosarcoma, small cell osteosarcoma, secondary osteosarcoma, and high-grade surface osteosarcoma.

Grading of conventional chondrosarcoma is based on cellularity, cytologic atypia, and mitotic figures, following the grading system proposed by Evans et al.⁴ Grade 1 (low-grade) chondrosarcoma is hypocellular and similar histologically to enchondroma. Grade 2 (intermediate-grade) chondrosarcoma is myxoid and more cellular/atypical than grade 1 chondrosarcoma. Grade 3 (high-grade) chondrosarcoma is hypercellular, pleomorphic, and contains observed mitotic activity.

Mesenchymal chondrosarcoma, fibrosarcoma, leiomyosarcoma, liposarcoma, undifferentiated pleomorphic sarcoma of bone and other “soft tissue-type” sarcomas that rarely occur in bone can be graded according to the French Federation of Cancer Centers Sarcoma Group (FNCLCC) grading system.⁵

Chordomas are locally aggressive lesions with a propensity for metastasis late in their clinical course and are not graded. Adamantinomas tend to have a low-grade clinical course, but this is variable. Fortunately, these are very rare. Other tumors such as periosteal chondrosarcoma (grading

does not predict behavior) or bone angiosarcoma (always considered high-grade behavior) are also not graded. According to the 2020 WHO classification of tumors of bone, adamantinomas are not graded.[2.3.6](#)

Bone Tumor Grades (Most Common)

Grade 1 (Low-Grade)

Low-grade intramedullary (central) osteosarcoma
Parosteal osteosarcoma
Grade I chondrosarcoma
Clear cell chondrosarcoma

Grade 2

Periosteal osteosarcoma
Grade II chondrosarcoma

Grade 3 (High-Grade)

Ewing sarcoma
Most round cell sarcomas
Sarcoma with BCOR genetic alterations
CIC-rearranged sarcoma
Conventional osteosarcoma
Telangiectactic osteosarcoma
Mesenchymal chondrosarcoma
Small cell osteosarcoma
Secondary osteosarcoma
High-grade surface osteosarcoma
Dedifferentiated chondrosarcoma
Dedifferentiated chordoma
Poorly differentiated chordoma
Malignancy in giant cell tumor (primary and secondary malignant giant cell tumor of bone)
Grade III chondrosarcoma
Leiomyosarcoma
Rhabdomyosarcoma
Undifferentiated pleomorphic sarcoma

TNM Grading

The 8th edition of the American Joint Committee on Cancer (AJCC) and International Union Against Cancer (UICC) staging system for bone tumors includes a 3-grade system but effectively collapses into high-grade and low-grade.[2.5](#) Other grading systems in (TNM) are based on differentiation, yet this is not applicable to primary intraosseous sarcomas.

GX	Grade cannot be assessed
G1	Well-differentiated, low-grade
G2	Moderately differentiated, high-grade
G3	Poorly differentiated, high-grade

For purposes of using the AJCC staging system (see note L), 3-grade systems can be converted to a 2-grade (TNM) system as follows: grade 1= low-grade; grade 2 and grade 3 = high-grade.

References

1. Inwards CY, Unni KK. Classification and grading of bone sarcomas. *Hematol Oncol Clin North Am.* 1995;9(3):545-569.
2. Amin MB, Edge SB, Greene FL, et al., eds. *AJCC Cancer Staging Manual.* 8th ed. New York, NY: Springer; 2017.
3. WHO Classification of Tumours Editorial Board. *Soft Tissue and Bone Tumors.* Lyon (France): International Agency for Research on Cancer; 2020. (WHO classification of tumours series, 5th ed.; vol. 3)
4. Evans HL, Ayala AG, Romsdahl MM. Prognostic factors in chondrosarcoma of bone: a clinicopathologic analysis with emphasis on histologic grading. *Cancer.* 1977 Aug;40(2):818-31.
5. Guillou L, Coindre JM, Bonichon F, et al. Comparative study of the National Cancer Institute and French Federation of Cancer Centers Sarcoma Group grading systems in a population of 410 adult patients with soft tissue sarcoma. *J Clin Oncol.* 1997;15(1):350-362.
6. Brierley JD, Gospodarowicz MK, Wittekind C, et al., eds. *TNM Classification of Malignant Tumours.* 8th ed. Oxford, UK: Wiley; 2016.

G. Mitotic Rate

Mitotic rate is determined by counting mitotic figures in the most mitotically active area, away from areas of necrosis, in either 10 consecutive high-power fields (HPF) (use the X40 objective) (1 HPF x 400 = 0.1734 mm²) or in the appropriate number of HPF to encompass 1 mm² based on each individual microscope.

The area of 1 HPF originally used measured 0.1734 mm². However, the area of 1 HPF using most modern microscopes with wider 40x lenses will most likely be higher. Pathologists are encouraged to determine the field area of their 40x lenses and divide 0.1734 by the obtained field area to obtain a conversion factor. The number of mitotic figures in 10 HPF multiplied by the obtained conversion factor and rounded to the nearest whole number should be used for reporting purposes.

An important change in the 5th Edition of the WHO Classification of Tumours series¹ is the conversion of mitotic count from the traditional denominator of 10 HPF to a defined area expressed in 1 mm², as an attempt to standardize the area used for mitotic count. Table 2 demonstrates the approximate number of fields required to encompass 1 mm² based on the field diameter and its corresponding area.

Table 2. Approximate number of fields per 1 mm² based on field diameter

Formula to calculate the area of one high power field of a specific microscope = $\frac{pr^2}{total\ magnification}$
Formula to calculate the field diameter = $\sqrt{\frac{pr^2}{total\ magnification}}$

Formula to calculate the field diameter = Objective Field Number/Objective Magnification

Field diameter (mm)	Area (mm ²)	Approximate number of fields per 1 mm ²
0.40	0.126	8
0.41	0.132	8
0.42	0.138	7
0.43	0.145	7

0.44	0.152	7
0.45	0.159	6
0.46	0.166	6
0.47	0.173	6
0.48	0.181	6
0.49	0.188	5
0.50	0.196	5
0.51	0.204	5
0.52	0.212	5
0.53	0.221	5
0.54	0.229	4
0.55	0.237	4
0.56	0.246	4
0.57	0.255	4
0.58	0.264	4
0.59	0.273	4
0.60	0.283	4
0.61	0.292	3
0.62	0.302	3
0.63	0.312	3
0.64	0.322	3
0.65	0.332	3
0.66	0.342	3
0.67	0.352	3
0.68	0.363	3
0.69	0.374	3

References

1. WHO Classification of Tumours Editorial Board. *Soft Tissue and Bone Tumors*. Lyon (France): International Agency for Research on Cancer; 2020. (WHO classification of tumours series, 5th ed.; vol. 3).

H. Response to Chemotherapy/Radiation Therapy Effect

It is essential to estimate neoadjuvant treatment effect in primary Ewing sarcoma and osteosarcoma of bone, as these have been shown to have prognostic significance.^{1,2,3,4,5,6} An entire representative slice of the tumor taken through the long axis should be mapped using a grid pattern diagram, photocopy, or radiologic film to indicate the site for each tumor block. In addition, the remainder of the neoplasm should be sampled at the rate of 1 section per centimeter. Areas of soft tissue extension and the interface of tumor with normal tissue should also be sampled. The sum of all viable areas measured microscopically is divided by the total cross-sectional area occupied by tumor to arrive at a percentage. One way to do so is to estimate the percent treatment effect (necrosis/fibrosis/hemorrhage/histiocytic response) on each slide of the grid slab (greatest surface area of tumor in the longest axis) and put these measurements into

an Excel spreadsheet and assess the average necrosis of all slides examined. This yields a finding that is compatible with treatment assessment. Prognostically significant therapy response in osteosarcoma, according to most series, is defined >90%, with those tumors showing >90% therapy response associated with a favorable prognosis.^{2,3,4} There are 2 protocols to assess response to therapy in Ewing sarcoma. Response can be assessed in the same manner as osteosarcoma or by the system of Picci, which is expressed as grade I (macroscopic viable tumor), grade II (microscopic viable tumor), or grade III (no viable tumor).^{5,6}

Histologic Classification of Treated Lesions

Due to extensive treatment effect, such as necrosis, fibrosis, and chemotherapy-induced and radiation-induced pleomorphism, it may not be possible to classify some lesions that were either never biopsied or where the biopsy was insufficient for a precise diagnosis. In problematic cases, the grade of the pretreatment specimen (i.e., biopsy, if available) should take precedence.

References

1. Amin MB, Edge SB, Greene FL, et al., eds. *AJCC Cancer Staging Manual*. 8th ed. New York, NY: Springer; 2017.
2. Picci P, Sangiorgi L, Rougraff BT, et al. Relationship of chemotherapy-induced necrosis and surgical margins to local recurrence in osteosarcoma. *J Clin Oncol*. 1994;12(12):2699-2705.
3. Raymond AK, Chawla SP, Carrasco CH, et al. Osteosarcoma chemotherapy effect: a prognostic factor. *Semin Diagn Pathol*. 1987;4(3):212-236.
4. Richardson SM, Wurtz LD, Collier CD. Ninety Percent or Greater Tumor Necrosis Is Associated With Survival and Social Determinants of Health in Patients With Osteosarcoma in the National Cancer Database. *Clin Orthop Relat Res*. 2023;481(3):512-522. PMID: 36099400; PMCID: PMC9928876.
5. Bacci G, Ferrari S, Bertoni F, et al. Prognostic factors in nonmetastatic Ewing's sarcoma of bone treated with adjuvant chemotherapy: analysis of 359 patients at the Istituto Ortopedico Rizzoli. *J Clin Oncol*. 2000;18(1):4-11.
6. Picci P, Bohling T, Bacci G, et al. Chemotherapy-induced tumor necrosis as a prognostic factor in localized Ewing's sarcoma of the extremities. *J Clin Oncol*. 1997;15(4):1553-1559.

I. Lymphatic and/or Vascular Invasion

Lymphatic or vascular invasion (LVI) indicates whether microscopic lymphatic or vascular invasion is identified. LVI includes lymphatic invasion or vascular invasion or both. By AJCC/UICC convention, LVI does not affect the T category indicating local extent of tumor unless specifically included in the definition of a T category.

J. Margins

It has been recommended that for all margins located less than 2 cm, the distance of the tumor from the margin be reported in centimeters.¹ However, there is a lack of agreement on this issue.² We recommend specifying the closest margin only and optionally the location of all margins less than 0.2 cm.³ Margins from bone tumors should be taken as *perpendicular* (radial) margins, if possible. If the tumor is located more than 2 cm from the bone margin, the marrow can be scooped out and submitted as a margin.

References

1. Abdul-Karim FW, Bauer TW, Kilpatrick SE, et al. Recommendations for the reporting of bone tumors. Association of Directors of Anatomic and Surgical Pathology. *Hum Pathol*. 2004;35(10):1173-1178.
2. Evrard R, Schubert T, Paul L, Docquier PL. Quality of resection margin with patient specific instrument for bone tumor resection. *J Bone Oncol*. 2022;34:100434. doi: 10.1016/j.jbo.2022.100434. PMID: 35601663; PMCID: PMC9115318.
3. Gomez-Brouchet A, Mascard E, Siegfried A, de Pinieux G, Gaspar N, Bouvier C, Aubert S, Marec-Bérard P, Piperno-Neumann S, Marie B, Larousserie F, Galant C, Fiorenza F, Anract P, Sales de Gauzy J, Gouin F; GROUPOS (GSF-GETO RESOS). Assessment of resection margins in bone sarcoma treated by neoadjuvant chemotherapy: Literature review and guidelines of the bone group (GROUPOS) of the French sarcoma group and bone tumor study group (GSF-GETO/RESOS). *Orthop Traumatol Surg Res*. 2019;105(4):773-780. Epub 2019. PMID: 30962172.

K. Regional Lymph Nodes

Regional lymph node metastasis is extremely rare in adult bone sarcomas. Nodes are not sampled routinely, and it is not necessary to exhaustively search for nodes. When no lymph nodes are resected, the pathologic 'N' category is not assigned (pNX is not used for bone tumors).¹ When present, regional lymph node metastasis has prognostic importance and should be reported. Patients whose nodal status is not determined to be positive for tumor, either clinically or pathologically, should be designated as N0.

References

1. Amin MB, Edge SB, Greene FL, et al, eds. *AJCC Cancer Staging Manual*. 8th ed. New York, NY: Springer; 2017.

L. pTNM Classification

The 8th edition TNM staging system for bone tumors of the AJCC and the UICC is recommended.^{1,2}

Anatomical staging using the AJCC system 8th ed. is considered clinically relevant only for the specific malignant entities listed in the World Health Organization classification 5th ed. This includes malignant chondrogenic tumors, osteogenic tumors, fibrogenic tumors, osteoclastic giant cell-rich tumors, notochordal tumors, myogenic tumors, lipogenic tumors, undifferentiated small round cell sarcomas and other mesenchymal tumors arising in bone. Locally aggressive entities such as synovial chondromatosis, osteoblastoma, giant cell tumor of bone, epithelioid hemangioma, pseudomyogenic hemangioma, and desmoplastic fibroma may be reported using this protocol but are not staged. Site groups for bone sarcomas are the following: appendicular skeleton, including trunk, skull and facial bones, pelvis, and spine. Pathologic staging includes pathologic data obtained from examination of a resected specimen sufficient to evaluate the highest T category, histopathologic type and grade, regional lymph nodes as appropriate, or distant metastasis. Because regional lymph node involvement from bone tumors is rare, the pathologic stage grouping includes any of the following combinations: pT pG pN pM, or pT pG cN cM, or cT cN pM. Biological grade should be assigned to all bone sarcomas, and based on published outcomes data, the current staging system accommodates a two-tiered (low- vs high-grade) system for recording grade. Histologic grading (G) uses a three-tiered system: G1 is considered low-grade, and G2 and G3 are grouped together as high-grade for biological grading.¹

TNM Descriptors

For identification of special cases of TNM or pTNM classifications, the “m” suffix and the “y” and “r” prefixes are used. Although they do not affect the stage grouping, they indicate cases needing separate analysis.

The “m” suffix indicates the presence of multiple primary tumors in a single site and is recorded in parentheses: pT(m)NM.

The “y” prefix indicates those cases in which classification is performed during or following initial multimodality therapy (i.e., neoadjuvant chemotherapy, radiation therapy, or both chemotherapy and radiation therapy). The cTNM or pTNM category is identified by a “y” prefix. The ycTNM or ypTNM categorizes the extent of tumor actually present at the time of that examination. The “y” categorization is not an estimate of tumor prior to multimodality therapy (i.e., before initiation of neoadjuvant therapy).

The “r” prefix indicates a recurrent tumor when staged after a documented disease-free interval and is identified by the “r” prefix: rTNM.

T Category Considerations (Figures 2 and 3)

Spine segments for staging:

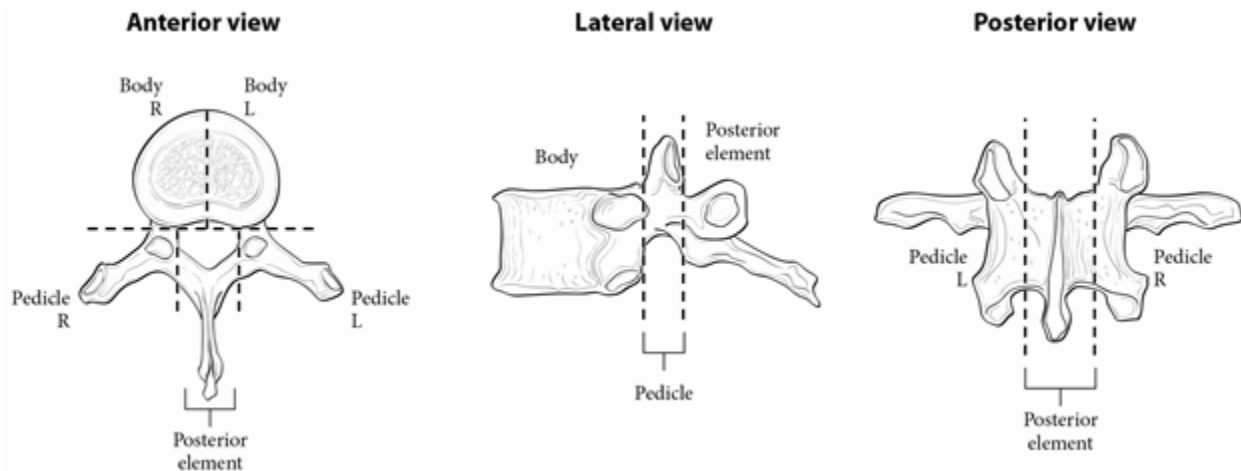


Figure 2. Spine segments for staging. Used with permission of the American Joint Committee on Cancer (AJCC), Chicago, Illinois. The original source for this material is the *AJCC Cancer Staging Manual* (2017) published by Springer Science and Business Media LLC, www.springerlink.com.

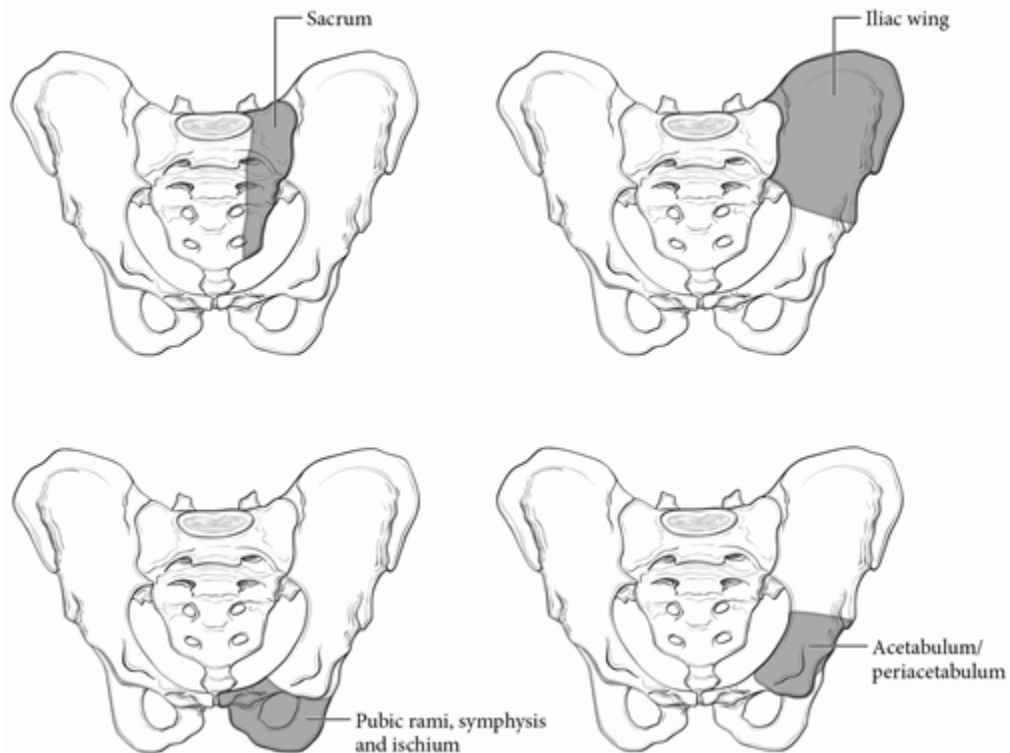


Figure 3. Pelvic segments for staging. Used with permission of the American Joint Committee on Cancer (AJCC), Chicago, Illinois. The original source for this material is the *AJCC Cancer Staging Manual* (2017) published by Springer Science and Business Media LLC, www.springerlink.com

N Category Considerations

Because of the rarity of lymph node involvement in sarcomas, the designation NX may not be appropriate and could be considered N0 if no clinical involvement is evident.

References

1. Amin MB, Edge SB, Greene FL, et al., eds. *AJCC Cancer Staging Manual*. 8th ed. New York, NY: Springer; 2017.
2. Brierley JD, Gospodarowicz MK, Wittekind C, et al., eds. *TNM Classification of Malignant Tumours*. 8th ed. Oxford, UK: Wiley; 2016.